

# Package: skater (via r-universe)

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**Title** Utilities for SNP-Based Kinship Analysis

**Description** Utilities for single nucleotide polymorphism (SNP) based kinship analysis testing and evaluation. The 'skater' package contains functions for importing, parsing, and analyzing pedigree data, performing relationship degree inference, benchmarking relationship degree classification, and summarizing identity by descent (IBD) segment data. Package functions and methods are described in Turner et al. (2021) ``skater: An R package for SNP-based Kinship Analysis, Testing, and Evaluation" <[doi:10.1101/2021.07.21.453083](https://doi.org/10.1101/2021.07.21.453083)>.

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**License** MIT + file LICENSE

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**BugReports** <https://github.com/signaturescience/skater/issues>

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arrange_ids	<i>Order IDs across two columns</i>
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### Description

Some types of data or results are indexed by two identifiers in two different columns corresponding to data points for *pairs* of observations. E.g., you may have columns called `id1` and `id2` that index the tibble for all possible pairs of results between samples A, B, and C. If you attempt to join two tibbles with `by=c("id1", "id2")`, the join will fail if samples are flipped from one dataset to another. E.g., one tibble may have `id1=A` and `id2=B` while the other has `id1=B` and `id2=A`. This function ensures that `id1` is alphanumerically first while `id2` is alphanumerically second. See examples.

### Usage

```
arrange_ids(.data, .id1, .id2)
```

### Arguments

<code>.data</code>	A tibble with two ID columns to arrange.
<code>.id1</code>	Unquoted name of the "id1" column. See examples.
<code>.id2</code>	Unquoted name of the "id2" column. See examples.

**Value**

A tibble with id1 and id2 rearranged alphanumerically.

**Examples**

```
d1 <- tibble::tribble(
  ~id1, ~id2, ~results1,
  "a", "b",      10L,
  "a", "c",      20L,
  "c", "b",      30L
)
d2 <- tibble::tribble(
  ~id1, ~id2, ~results2,
  "b", "a",     101L,
  "c", "a",     201L,
  "b", "c",     301L
)
# Inner join fails because id1!=id2.
dplyr::inner_join(d1, d2, by=c("id1", "id2"))
# Arrange IDs
d1 %>% arrange_ids(id1, id2)
d2 %>% arrange_ids(id1, id2)
# Inner join
dplyr::inner_join(arrange_ids(d1, id1, id2), arrange_ids(d2, id1, id2), by=c("id1", "id2"))
# Recursively, if you had more than two tibbles
list(d1, d2) %>%
  purrr::map(arrange_ids, id1, id2) %>%
  purrr::reduce(dplyr::inner_join, by=c("id1", "id2"))
```

---

 calc\_accuracy

*Calculate Accuracy*


---

**Description**

Calculates accuracy and related metrics.

**Usage**

```
calc_accuracy(table)
```

**Arguments**

table            A frequency table created with [table](#)

**Details**

Calculates accuracy, lower and upper bounds, the guessing rate and p-value of the accuracy vs. the guessing rate. This function is called by `confusion_matrix`, but if this is all you want, you can simply supply the table to this function.

**Value**

A tibble with the corresponding statistics

**Author(s)**

Michael Clark (see [m-clark/confusion\\_matrix](#)).

**See Also**

[binom.test](#)

---

calc\_stats

*Calculate various statistics from a confusion matrix*

---

**Description**

Given a frequency table of predictions versus target values, calculate numerous statistics of interest.

**Usage**

```
calc_stats(tabble, prevalence = NULL, positive, ...)
```

**Arguments**

tabble	A frequency table created with <a href="#">table</a>
prevalence	Prevalence value. Default is NULL
positive	Positive class
...	Other, not currently used

**Details**

Used within `confusion_matrix` to calculate various confusion matrix metrics. This is called by `confusion_matrix`, but if this is all you want you can simply supply the table.

Suppose a 2x2 table with notation

	target	
Predicted	Event	No Event
Event	A	B
No Event	C	D

The formulas used here are:

$$\text{Sensitivity} = A / (A + C)$$

$$\text{Specificity} = D / (B + D)$$

$$\text{Prevalence} = (A + C) / (A + B + C + D)$$

$$\text{PositivePredictiveValue} = (\text{sensitivity} * \text{prevalence}) / ((\text{sensitivity} * \text{prevalence}) + ((1 - \text{specificity}) * (1 - \text{prevalence})))$$

$$\text{NegativePredictiveValue} = (\text{specificity} * (1 - \text{prevalence})) / (((1 - \text{sensitivity}) * \text{prevalence}) + ((\text{specificity}) * (1 - \text{prevalence})))$$

$$\text{DetectionRate} = A / (A + B + C + D)$$

$$\text{DetectionPrevalence} = (A + B) / (A + B + C + D)$$

$$\text{BalancedAccuracy} = (\text{sensitivity} + \text{specificity}) / 2$$

$$\text{Precision} = A / (A + B)$$

$$\text{Recall} = A / (A + C)$$

$$F1 = \text{harmonicmeanofprecisionandrecall} = (1 + \text{beta}^2) * \text{precision} * \text{recall} / ((\text{beta}^2 * \text{precision}) + \text{recall})$$

where beta = 1 for this function.

$$\text{FalseDiscoveryRate} = 1 - \text{PositivePredictiveValue}$$

$$\text{FalseOmissionRate} = 1 - \text{NegativePredictiveValue}$$

$$\text{FalsePositiveRate} = 1 - \text{Specificity}$$

$$\text{FalseNegativeRate} = 1 - \text{Sensitivity}$$

$$D' = qnorm(\text{Sensitivity}) - qnorm(1 - \text{Specificity})$$

$$AUC = pnorm(D' / \text{sqrt}(2))$$

See the references for discussions of the first five formulas. Abbreviations:

**Positive Predictive Value: PPV**

**Negative Predictive Value: NPV**

**False Discovery Rate: FDR**

**False Omission Rate: FOR**

**False Positive Rate: FPR**

**False Negative Rate: FNR**

## Value

A tibble with (at present) columns for sensitivity, specificity, PPV, NPV, F1 score, detection rate, detection prevalence, balanced accuracy, FDR, FOR, FPR, FNR. For more than 2 classes, these statistics are provided for each class.

## Note

Different names are used for the same statistics.

**Sensitivity: True Positive Rate, Recall, Hit Rate, Power**

**Specificity: True Negative Rate**

**Positive Predictive Value: Precision**

**False Negative Rate: Miss Rate, Type II error rate, beta**

**False Positive Rate: Fallout, Type I error rate, alpha**

This function is called by `confusion_matrix`, but if this is all you want, you can simply supply the table to this function.

**Author(s)**

Michael Clark (see [m-clark/confusion\\_matrix](#)).

**References**

Kuhn, M. (2008), "Building predictive models in R using the caret package," *Journal of Statistical Software*, (<https://www.jstatsoft.org/article/view/v028i05>).

Altman, D.G., Bland, J.M. (1994) "Diagnostic tests 1: sensitivity and specificity", *British Medical Journal*, vol 308, 1552.

Altman, D.G., Bland, J.M. (1994) "Diagnostic tests 2: predictive values," *British Medical Journal*, vol 309, 102.

Velez, D.R., et. al. (2008) "A balanced accuracy function for epistasis modeling in imbalanced datasets using multifactor dimensionality reduction.," *Genetic Epidemiology*, vol 4, 306.

---

confusion\_matrix

*Calculate various statistics from a confusion matrix*

---

**Description**

Given a vector of predictions and target values, calculate numerous statistics of interest. Modified from [m-clark/confusion\\_matrix](#).

**Usage**

```
confusion_matrix(
  prediction,
  target,
  positive = NULL,
  prevalence = NULL,
  dnn = c("Predicted", "Target"),
  longer = FALSE,
  ...
)
```

**Arguments**

prediction	A vector of predictions
target	A vector of target values
positive	The positive class for a 2-class setting. Default is NULL, which will result in using the first level of target.
prevalence	Prevalence rate. Default is NULL.
dnn	The row and column headers for the contingency table returned. Default is 'Predicted' for rows and 'Target' for columns.
longer	Transpose the output to long form. Default is FALSE (requires tidy 1.0).
...	Other parameters, not currently used.

**Details**

This returns accuracy, agreement, and other statistics. See the functions below to find out more. Originally inspired by the confusionMatrix function from the caret package.

**Value**

A list of tibble(s) with the associated statistics and possibly the frequency table as list column of the first element. If classes contain >1 numeric class and a single non-numeric class (e.g., "1", "2", "3", and "Unrelated", the RMSE of the reciprocal of the Targets + 0.5 will also be returned.)

**References**

Kuhn, M., & Johnson, K. (2013). Applied predictive modeling.

**See Also**

[calc\\_accuracy](#) [calc\\_stats](#)

**Examples**

```
prediction = c(0,1,1,0,0,1,0,1,1,1)
target     = c(0,1,1,1,0,1,0,1,0,1)
confusion_matrix(prediction, target, positive = '1')

set.seed(42)
prediction = sample(letters[1:4], 250, replace = TRUE, prob = 1:4)
target     = sample(letters[1:4], 250, replace = TRUE, prob = 1:4)
confusion_matrix(prediction, target)

prediction = c(rep(1, 50), rep(2, 40), rep(3, 60))
target     = c(rep(1, 50), rep(2, 50), rep(3, 50))
confusion_matrix(prediction, target)
confusion_matrix(prediction, target) %>% purrr::pluck("Table")
confusion_matrix(prediction, target, longer=TRUE)
confusion_matrix(prediction, target, longer=TRUE) %>%
  purrr::pluck("Other") %>%
  tidyr::spread(Class, Value)

# Prediction with an unrelated class
prediction = c(rep(1, 50), rep(2, 40), rep(3, 60), rep("Unrelated", 55))
target     = c(rep(1, 50), rep(2, 50), rep(3, 55), rep("Unrelated", 50))
confusion_matrix(prediction, target)
# Prediction with two unrelated classes
prediction = c(rep(1, 50), rep(2, 40), rep("Third", 60), rep("Unrelated", 55))
target     = c(rep(1, 50), rep(2, 50), rep("Third", 55), rep("Unrelated", 50))
confusion_matrix(prediction, target)
```

---

dibble	<i>Degree tibble</i>
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---

**Description**

Creates a tibble with degree, expected kinship coefficient, and inference boundaries.

Rows will be created up to the `max_degree`, with an additional row for any relationship more distant than `max_degree`. The degree value for the final row will be NA. This represents inference criteria for "unrelated" individuals. See examples.

**Usage**

```
dibble(max_degree = 3L)
```

**Arguments**

`max_degree`      The most distant degree you want to measure (usually between 3-9, default 3).

**Value**

A tibble containing the degree, expected kinship coefficient (k), lower (l) and upper (u) inference bounds.

**Examples**

```
dibble(3)
dibble(10)
```

---

fam2ped	<i>Fam to pedigree</i>
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---

**Description**

Converts a **PLINK-formatted fam file** to a pedigree object using `kinship2::pedigree`.

**Usage**

```
fam2ped(fam)
```

**Arguments**

`fam`                      A tibble with six columns of PLINK .fam data as read in by `read_fam`.

**Value**

A tibble with new listcol ped containing pedigrees from `kinship2::pedigree`.



## Examples

```
famfile <- system.file("extdata", "3gens.fam", package="skater", mustWork=TRUE)
fam <- read_fam(famfile)
fam2ped(fam)
```

---

ibd2kin	<i>Compute kinship coefficient from IBD segments</i>
---------	--

---

## Description

This function is used to retrieve a relatedness measure from IBD segments. The relatedness value returned is the kinship coefficient.

## Usage

```
ibd2kin(.ibd_data, .map, type = NULL)
```

## Arguments

<code>.ibd_data</code>	Tibble with IBD segments created using the <a href="#">read_ibd</a> function
<code>.map</code>	Tibble with the genetic map data created using the <a href="#">read_map</a> function
<code>type</code>	Type of IBD to use for kinship coefficient calculation; must be 'IBD1', 'IBD2', or NULL (both IBD1 and IBD2 will be treated the same); default is NULL

## Details

The input data should be pairwise IBD segments prepared via [read\\_ibd](#). The function will internally loop over each chromosome, and use a specified genetic map to convert shared segments to genetic units. After doing so, the function converts the shared length to a kinship coefficient by summing  $0.5 * IBD2 + 0.25 * IBD1$ .

Note that the data read in by [read\\_ibd](#) when `source="pedsim"` returns a list with separate tibbles for IBD1 and IBD2 segments. The current implementation of this function requires running this function independently on IBD1 and IBD2 segments, then summarizing (adding) the corresponding proportions. See examples.

## Value

Tibble with three columns:

1. id1 (sample identifier 1)
2. id2 (sample identifier 2)
3. kinship (kinship coefficient derived from shared segments)

## References

[http://faculty.washington.edu/sguy/ibd\\_relatedness.html](http://faculty.washington.edu/sguy/ibd_relatedness.html)

## Examples

```
pedsim_fp <- system.file("extdata", "GBR.sim.seg.gz", package="skater", mustWork=TRUE)
pedsim_seg <- read_ibd(pedsim_fp, source = "pedsim")
gmapfile <- system.file("extdata", "sexspec-avg-min.plink.map", package="skater", mustWork=TRUE)
gmap <- read_map(gmapfile)
ibd1_dat <- ibd2kin(.ibd_data=pedsim_seg$IBD1, .map=gmap, type="IBD1")
ibd2_dat <- ibd2kin(.ibd_data=pedsim_seg$IBD2, .map=gmap, type="IBD2")
dplyr::bind_rows(ibd1_dat,ibd2_dat) %>%
  dplyr::group_by(id1,id2) %>%
  dplyr::summarise(kinship = sum(kinship), .groups = "drop")
```

---

interpolate

*Interpolate over segments*

---

## Description

This is an unexported helper used in `ibd2kin`. The function interpolates over segments to apply genetic length to the segment. It is inspired by Python code distributed by the Browning lab ([documentation](#)).

## Usage

```
interpolate(ibd_bp, chromgpos)
```

## Arguments

<code>ibd_bp</code>	Base pair for the IBD segment over which to interpolate
<code>chromgpos</code>	Genetic map data for a specific chromosome

## Value

Numeric vector with the genetic distance shared at the segment.

## References

[http://faculty.washington.edu/sguy/ibd\\_relatedness.html](http://faculty.washington.edu/sguy/ibd_relatedness.html)

---

kin2cm	<i>Kinship coefficient to cM</i>
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---

### Description

"Converts" a kinship coefficient to put on the same scale as shared cM using the formula  $cm < -pmin(3560, 4 * pmax(0, k) * 3560)$ .

### Usage

```
kin2cm(k)
```

### Arguments

k	Kinship coefficient (numeric, typically between 0 and .5, although KING can produce values <0).
---	---

### Value

A vector of numeric estimated cM, ranging from 0-3560.

### References

<https://dnainter.com/tools/sharedcmv4>.

<https://www.ancestry.com/dna/resource/whitePaper/AncestryDNA-Matching-White-Paper.pdf>.

<https://verogen.com/wp-content/uploads/2021/03/snp-typing-uas-kinship-estimation-gedmatch-pro-tech.pdf>.

### Examples

```
kin2cm(.25)
kin2cm(.125)
kin2cm(.0625)
dibble(9) %>% dplyr::mutate(cm=kin2cm(k))
```

---

kin2degree	<i>Kinship coefficient to degree</i>
------------	--------------------------------------

---

**Description**

Infers relationship degree given a kinship coefficient.

**Usage**

```
kin2degree(k, max_degree = 3L)
```

**Arguments**

k	Kinship coefficient (numeric, typically between 0 and .5, although KING can produce values <0).
max_degree	Max degree resolution (default 3). Used to seed <a href="#">dibble</a> . Anything below the inference range of max_degree will report NA. See <a href="#">dibble</a> .

**Value**

A vector with inferred degree, up to the maximum degree in [dibble](#) (anything more distant is NA, i.e., unrelated).

**Examples**

```
kin2degree(0.5)
kin2degree(0.25)
kin2degree(0.125)
kin2degree(0.0625)
kin2degree(0.03125)
kin2degree(0.03125, max_degree=5)
kin2degree(-0.05)
k <- seq(.02, .5, .03)
kin2degree(k)
kin2degree(k, max_degree=5)
tibble::tibble(k=k) %>% dplyr::mutate(degree=kin2degree(k))
```

---

ped2kinpair	<i>Pedigree to pairwise kinship</i>
-------------	-------------------------------------

---

**Description**

Converts a pedigree class object from [fam2ped](#) to a pairwise list of relationships and their expected/theoretical kinship coefficient.

**Usage**

```
ped2kinpair(ped)
```

**Arguments**

ped                    A "pedigree" class object from [fam2ped](#).

**Value**

A tibble containing all pairwise kinship coefficients from the input pedigree.

**Examples**

```
famfile <- system.file("extdata", "3gens.fam", package="skater", mustWork=TRUE)
famfile %>%
  read_fam() %>%
  fam2ped() %>%
  dplyr::mutate(kinpairs=purrr::map(ped, ped2kinpair)) %>%
  dplyr::select(fid, kinpairs) %>%
  tidyr::unnest(cols=kinpairs)
```

---

plot\_pedigree

*Plot pedigree*

---

**Description**

Plot pedigree

**Usage**

```
plot_pedigree(ped, file = NULL, width = 10, height = 8)
```

**Arguments**

ped                    List of pedigree objects from [fam2ped](#)  
file                    Output file path (must end in ".pdf")  
width                   Width of output PDF  
height                   Height of output PDF

**Value**

No return value, called for side effects.

---

read_akt	<i>Read AKT kin output file</i>
----------	---------------------------------

---

### Description

Reads in an akt kin **results file**. Input file must have seven columns, whitespace delimited:

1. id1 (member 1)
2. id2 (member 2)
3. IBD0 (ratio of IBD0/All SNPS)
4. IBD1 (ratio of IBD1/All SNPS)
5. Kinship Coefficient
6. NSNPS

### Usage

```
read_akt(file)
```

### Arguments

file	Input file path
------	-----------------

### Value

A tibble containing the 7 columns from the akt file.

### Examples

```
aktFile <- system.file("extdata", "3gens.akt", package="skater", mustWork=TRUE)
akt <- read_akt(aktFile)
akt
```

---

read_fam	<i>Read PLINK-formatted .fam file</i>
----------	---------------------------------------

---

### Description

Reads in a **PLINK-formatted .fam file**. Input file must have six columns:

1. Family ID
2. Individual ID
3. Father ID
4. Mother ID
5. Sex
6. Affected Status

**Usage**

```
read_fam(file)
```

**Arguments**

file                    Input file path

**Value**

A tibble containing the 6 columns from the fam file.

**Examples**

```
famfile <- system.file("extdata", "3gens.fam", package="skater", mustWork=TRUE)
fam <- read_fam(famfile)
fam
```

---

read_ibd	<i>Read IBD segment file</i>
----------	------------------------------

---

**Description**

Reads in the inferred IBD segments from hapibd ([documentation](#)) or IBD segment file generated by ped-sim ([documentation](#)).

If reading a hapibd segment file, the input data should have the following columns:

1. First sample identifier
2. First sample haplotype index (1 or 2)
3. Second sample identifier
4. Second sample haplotype index (1 or 2)
5. Chromosome
6. Base coordinate of first marker in segment
7. Base coordinate of last marker in segment
8. cM length of IBD segment

If read a pedsim segment file, the input data should have the following columns:

1. First sample identifier
2. Second sample identifier
3. Chromosome
4. Physical position start
5. Physical position end
6. IBD type
7. Genetic position start
8. Genetic position end
9. Genetic length (end - start)

**Usage**

```
read_ibd(file, source)
```

**Arguments**

file	Input file path
source	Source of the input file; must be one of "hapibd" or "pedsim"

**Value**

if source="hapibd", a tibble is returned. If source="pedsim", a list with two tibble elements, IBD1 and IBD2 is returned. Both the hapibd tibble, and the two pedsim tibbles contain six columns:

1. id1 (sample identifier 1)
2. id2 (sample identifier 2)
3. chr (chromosome)
4. start (segment bp start coordinate)
5. end (segment bp end coordinate)
6. length (shared segment length in genetic units, cM)

**References**

<https://github.com/browning-lab/hap-ibd#output-files>

<https://github.com/williamslab/ped-sim#output-ibd-segments-file>

**Examples**

```
hapibd_fp <- system.file("extdata", "GBR.sim.ibd.gz", package="skater", mustWork=TRUE)
hapibd_seg <- read_ibd(hapibd_fp, source = "hapibd")
pedsim_fp <- system.file("extdata", "GBR.sim.seg.gz", package="skater", mustWork=TRUE)
pedsim_seg <- read_ibd(pedsim_fp, source = "pedsim")
```

---

read\_ibis

*Read IBIS coef output file*

---

**Description**

Reads in an ibis **results file**. Input file must have six columns, whitespace delimited:

1. id1 (member 1)
2. id2 (member 2)
3. Kinship Coefficient
4. IBD2 (ratio of IBD2/All SNPS)
5. Segment count
6. Kinship Degree



**Usage**

```
read_ibis(file)
```

**Arguments**

file            Input file path

**Value**

A tibble containing the 6 columns from the ibis file.

**Examples**

```
ibisFile <- system.file("extdata", "3gens.ibis.coef", package="skater", mustWork=TRUE)
ibis <- read_ibis(ibisFile)
ibis
```

---

read_map	<i>Read genetic map file</i>
----------	------------------------------

---

**Description**

This function reads in the content from a genetic map file to translate physical distance to genetic units (i.e. cM). Regardless of the source, the input file must be sex-averaged and in a tab-separated "Plink" format ([documentation](#)) with the following four columns and no header (i.e. no column names):

1. Chromosome
2. Identifier (ignored in read\_map())
3. Length (genetic length within the physical position boundary)
4. Position (physical position boundary)

The columns must be in the order above. Note that only the first, third, and fourth columns are used in the function.

**Usage**

```
read_map(file)
```

**Arguments**

file            Input file path

## Details

The genetic map could come from different sources. One source is the HapMap map distributed by the Browning Lab ([documentation](#)). If this map file is used, the non-sex chromosomes can be downloaded and concatenated to a single file as follows:

```
wget https://bochet.gcc.biostat.washington.edu/beagle/genetic_maps/plink.GRCh37.map.zip
unzip plink.GRCh37.map.zip
cat *chr[0-9]*GRCh37.map | sort -k1,1 -k4,4 --numeric-sort > plink.allchr.GRCh37.map
```

Another source is a sex-specific map ("bherer") originally published by Bherer et al and recommended by the developers of ped-sim for simulating IBD segments ([documentation](#)). To retrieve and prep this map file for simulation:

```
# Get the refined genetic map and extract
wget --no-check-certificate https://github.com/cbherer/Bherer_etal_SexualDimorphismRecombination/raw
tar xvpz Refined_genetic_map_b37.tar.gz

# Format for ped-sim as per https://github.com/williamslab/ped-sim#map-file-
printf "#chr\tpos\tmale_cM\tfemale_cM\n" > sexspec.pedsim.map
for chr in {1..22}; do
  paste Refined_genetic_map_b37/male_chr$chr.txt Refined_genetic_map_b37/female_chr$chr.txt \
    | awk -v OFS="\t" 'NR > 1 && $2 == $6 {print $1,$2,$4,$8}' \
    | sed 's/^chr//' >> sexspec.pedsim.map;
done

# Clean up
rm -rf Refined_genetic_map_b37*
```

After this, the sexspec.pedsim.map file is ready for use in simulation. However, it must be averaged and reformatted to "Plink format" to use here:

```
cat sexspec.pedsim.map | grep -v "^#" | awk -v OFS="\t" '{print $1, ".", ($3+$4)/2, $2}' > sexspec-avg.plin
```

# The genetic maps created above are in the tens of megabytes size range. This is trivial to store for most systems but a reduced version would increase portability and ease testing. This "minimum viable genetic map" could be used for testing and as installed package data in an R package for example analysis. Read more about minimum viable genetic maps at:

- Blog post: <https://hapi-dna.org/2020/11/minimal-viable-genetic-maps/>
- Github repo with python code: [https://github.com/williamslab/min\\_map](https://github.com/williamslab/min_map)

The code as written below reduces the averaged sex-specific genetic map from 833776 to 28726 positions (~30X reduction!).

```
# Get minmap script from github
wget https://raw.githubusercontent.com/williamslab/min_map/main/min_map.py

# Create empty minmap
```

```

echo -n > sexspec-avg-min.plink.map

# For each autosome...
for chr in {1..22}; do
  echo "Working on chromosome $chr..."
  # First pull out just one chromosome
  grep "^${chr}[[[:space:]]" sexspec-avg.plink.map > tmp.${chr}
  # Run the python script on that chromosome.
  # The genetic map column is 3rd column (2nd in 0-start). Physical position is last column (3 in 0-based)
  python3 min_map.py -mapfile tmp.${chr} -chr ${chr} -genetcol 2 -physcol 3 -noheader -error 0.05
  # Strip out the header and reformat back to plink format, and append to minmap file
  cat min_viable_map${chr}.txt | grep -v "^#" | awk -v OFS="\t" '{print $1, ".", $4, $2}' >> sexspec-avg-min
  # Clean up
  rm -f min_viable_map${chr}.txt tmp.${chr}
done

```

This averaged version of the Bherer sex-specific map, reduced to a minimum viable genetic map with at most 5% error, in Plink format, is available as installed package data (see examples). This is useful for testing code, but the full genetic map should be used for most analysis operations.

### Value

A tibble containing 3 columns:

1. chr (chromosome)
2. value (genetic length within the physical position boundary)
3. bp (physical position boundary)

### References

<https://www.cog-genomics.org/plink/1.9/formats#map>  
[https://bochet.gcc.biostat.washington.edu/beagle/genetic\\_maps/](https://bochet.gcc.biostat.washington.edu/beagle/genetic_maps/)  
<https://github.com/williamslab/ped-sim#map-file>  
<https://www.nature.com/articles/ncomms14994>  
<https://www.nature.com/articles/ncomms14994>  
[https://github.com/cbherer/Bherer\\_etal\\_SexualDimorphismRecombination](https://github.com/cbherer/Bherer_etal_SexualDimorphismRecombination)

### Examples

```

gmapfile <- system.file("extdata", "sexspec-avg-min.plink.map", package="skater", mustWork=TRUE)
gmap <- read_map(gmapfile)

```

---

read_plink2_king	<i>Read PLINK KING table</i>
------------------	------------------------------

---

### Description

Reads in the output from `plink2 --make-king-table` ([documentation](#)). Input file must have six columns, tab delimited:

1. id1 (member 1)
2. id2 (member 2)
3. nsnps
4. hethet: proportion of sites where both are heterozygous
5. k: Kinship Coefficient

### Usage

```
read_plink2_king(file)
```

### Arguments

file	Input file path
------	-----------------

### Value

A tibble containing the 6 columns from the `plink2 --make-king-table` output.

### References

[https://www.cog-genomics.org/plink/2.0/distance#make\\_king](https://www.cog-genomics.org/plink/2.0/distance#make_king)

### Examples

```
plink2kingFile <- system.file("extdata", "plink2-king-table.tsv", package="skater", mustWork=TRUE)
plink2king <- read_plink2_king(plink2kingFile)
plink2king
plink2king %>% dplyr::filter(k>0.01)
```

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